Synthesis of Fluorinated β -Aminophosphonates and γ -Lactams

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S Supporting Information

[AB](#page-7-0)STRACT: [The functio](#page-7-0)nalized polyfluorophosphorylated 1-azadienes I have been prepared by a Wittig reaction of ethyl glyoxalate and perfluorophosphorylated conjugated phosphoranes, obtained by reaction of phosphazenes and fluorinated acetylenic phosphonates. Subsequent reduction of both carbon− carbon and carbon−nitrogen double bonds of these 1-azadienes

I affords the fluorine-containing β-aminophosphonates II, with the syn β-aminophosphonate being obtained as the major diastereoisomer. Base-mediated cyclocondensation of a diastereomeric mixture of aminophosphonates II leads exclusively to a new type of functionalized trans-γ-lactams III in a diastereoselective way. A computational study has also been used to explain the observed diastereoselectivity of these reactions.

■ INTRODUCTION

Organophosphorus compounds are important substrates in the study of biochemical processes,¹ and β -aminophosphonates, as they are isosteres of β -amino acids, play an important role as enzyme inhibitors, agrochemicals, and [p](#page-7-0)harmaceuticals² as well as reveal diverse and interesting biological and biochemical properties. Some β -aminop[h](#page-7-0)osphonic acids and their derivatives have been identified as natural products.^{3a,b} The parent acid was first isolated from Celiata protozoa, $3c$ and subsequently the compound along with its different derivative[s w](#page-7-0)as obtained from microorganisms.^{3d,e} O[t](#page-7-0)her important uses of these compounds are as pesticides 3^f and in the preparation of valuable metal complexes.^{3g,h}

Furthermore, the introduction of a fluorine atom or a fluorinat[ed](#page-7-0) moiety can modulate the properties of a bioactive m[olec](#page-7-0)ule, since this may lead to changes in solubility, lipophilicity, metabolic stability, conformation, hydrogen-bonding ability, or chemical reactivity.⁴ Due to the unique properties of the fluorine atom, fluorinated molecules occupy a significant place⁵ in pharmaceut[ic](#page-7-0)al/medicinal,⁶ agrochemical,⁷ and materials sciences.⁸ Fluorine incorporation into a chemical structure ha[s](#page-7-0) been used in pharmaceutic[al](#page-7-0) developme[nt](#page-7-0) and drug design t[o](#page-7-0) prevent molecules from being metabolized too quickly, thereby allowing a drug to act before it is cleared from the body.⁹ In this respect, particular interest has focused on developing synthetic methods for the preparation of fluorinated building blocks, $6,10$ among them fluorinated aminophosphonates.¹¹ Efficient examples of aminophosphonates containing fluoroalkyl groups as liga[nds](#page-7-0) for phosphoglycerate kinase^{12a} or antibacterials,^{12b} inhibitors of serine esterases, alanine racemase, and pyrimidine phosphorylases, $12c$ and also the preparation of [fl](#page-7-0)uorinated peptid[omi](#page-7-0)metics¹³ have been demonstrated. However, the literature contains very [few](#page-7-0) preparative methods for the preparation of fluorinated β -aminophosphonates, such as by the addition of amines to unsaturated phosphonates,^{12b,14} by the addition of fluorinated phosphonate carbanions to

N-protected haloamines^{12a} or imines,¹⁵ or by ring opening of fluoroalkyl aziridine-2-phosphonates.¹⁶

On the other hand, [γ](#page-7-0)-lactams h[ave](#page-7-0) important applications in the drug-discovery process, as [ke](#page-7-0)y intermediates in the preparation of biologically and pharmaceutically relevant molecules in the treatment of cancer,^{17a} fungal infections,^{17b} epilepsy,^{17c,d} HIV ,^{17e,f} neurodegenerative diseases,^{17g} and depression.^{17h} The *γ*-lactam core showed a bett[er](#page-7-0) fit than [the](#page-8-0) larger δ -lactam [core](#page-8-0) into the narrow hydrophobic pocket [of](#page-8-0) histone deacetyla[se \(](#page-8-0)HDAC) active site and showed better inhibition profiles of HDACs and cancer cell growth inhibitory activities.^{18a} γ -Lactams are also viable glycinamide replacements within a series of cyclohexane-based CCR2 antagonists which could fi[nd](#page-8-0) additional use in the design and development of future antagonists.^{18b}

Continuing with our interest in the chemistry of fluorinated aminophosphorus derivatives as well as of [elec](#page-8-0)tron-rich, 19 fluorinated, 20 and functionalized 1-azadienes, 21 here we report the diastereolective preparation of novel fluoroalkyl- and phosph[oru](#page-8-0)s-substitut[ed](#page-8-0) γ-lactams I and fluorine-cont[ain](#page-8-0)ing $β$ -aminophosphonates II from functionalized 1-azadienes III, easily prepared from phosphazenes IV, polyfluoroacetylenephosphonates V, and ethyl glyoxalate VI (Scheme 1).

■ RES[U](#page-1-0)LTS AND DISCUSSION

The formation of conjugated phosphoranes by the reaction of phosphazenes and acetylenic esters has been reported.²² Therefore, a convenient method for the synthesis of some diethoxyphosphinyl perfluoroalkylidene phosphoranes based [on](#page-8-0) a $\lceil 2 + 2 \rceil$ cycloaddition reaction between phosphazenes 1 and polyfluoroacetylenephosphonates 2 has been developed (Scheme 2).

Received: February 6, 2013 Published: March 13, 2013

Scheme 1

Phosphazenes 1 were readily prepared by addition of the corresponding phosphines to azides through the Staudinger reaction.²³ Given the instability of P-alkylphosphazene species, 24 they were allowed to react in situ with polyfluoroacetylenephosphonates [2](#page-8-0) prepared according to the established procedure.²⁵ Rea[cti](#page-8-0)on progress for the formation of ylides 4 was monitored by $31P$ NMR spectroscopy. Signals for the phosphazenes (δ 2[−](#page-8-0)10 ppm) and the yne phosphonates (δ −9 to −11 ppm)²⁵ disappear, and resonances corresponding to polyfluorophosphorylated ylides 4 appear in the range δ 15−26 ppm. ¹⁹F NMR could [be](#page-8-0) applied similarly, with the loss of 2 (δ −53 to −112 ppm) and the appearance of 4 (δ −60 to −70 ppm). The results are summarized in Table 1.

The structure of isolated ylide $4g (R = Ph)$ was determined by NMR spectroscopy and mass spectrometry. The ³¹P NMR spectrum of ylide 4g presents two doublets for phosphonate and phosphorane phosphorus atoms, observed respectively at δ 24.2 and 25.7 ppm with the coupling constant ²J_{pp} = 51.1 Hz. The formation of phosphoranes 4 could be explained by $[2 + 2]$ cycloaddition between phosphazenes 1 and polyfluoroacetylenephosphonates 2, via intermediates 3 followed by ring opening, affording polyfluoroalkylated diethoxyphosphinyl phosphoranes 4 (Scheme 2) in a way similar to that reported for phosphazenes and phosphorus ylides with acetylenic esters.²² With the exception of trifluoromethylphosphorylated ylide $4g (R = Ph)$, which could be isolated and purified by recrystallizatio[n](#page-8-0) in hexane (68% yield, Table 1, entry 7), in general polyfluorophosphorylated ylides 4 were unstable and were used in situ without purification in further reactions. In fact, water-assisted hydrolysis of trifluoromethyl diethoxyphosphorylated ylide 4a ($R_F = CF_3$, $Ar = p$ -MeOC₆H₄) rapidly yielded the corresponding hydrolyzed product as a mixture of imine-enamine tautomers 5/5′ (Scheme 2).

Subsequently, the Wittig reaction of ylides 4 with carbonyl compounds was explored. The perfluoroalkylated conjugated

Table 1. Polyfluorophosphorylated Ylides 4 Obtained

		entry compd	PR ₃	R_F	Ar	reaction conditions	
	1	4a	PMe ₃	CF ₃	p -MeO-C ₆ H ₄	$CHCl3/30$ min/room temp	
	\mathfrak{p}	4b	PMe ₃	CF ₃ p -Me-C ₆ H ₄ temp		CHCl ₃ /30 min/room	
	3	4c	PMe ₃	CF ₃	p -NO ₂ -C ₆ H ₄	$CHCl3/30$ min/room temp	
	$\overline{4}$	4d	PMe_2Ph	CF ₃	p -MeO-C ₆ H ₄	CHCl ₃ /30 min/room temp	
	5	4e	PMe_2Ph	CF ₃	p -Me-C ₆ H ₄	$CHCl3/1$ h/room temp	
	6	4f	PMe_2Ph	CF ₃	$p\text{-NO}_2\text{-C}_6\text{H}_4$	$CHCl3/15$ h/room temp CHCl ₃ /21.5 h/ Δ	
	7	$4g^a$	PPh ₃	CF ₃	p -Me-C ₆ H ₄		
						toluene/17.5h/ Δ	
	8	4h	PMe ₃	C_2F_5	p -MeO-C ₆ H ₄	$CHCl3/3$ h/room temp	
	9	4i	PMe ₃	C_2F_5	$p\text{-NO}_2\text{-C}_6\text{H}_4$	CHCl ₃ /30 h/ Δ	
	10	4j	PMe ₃	CF ₂ H	p -NO ₂ -C ₆ H ₄	toluene/4 h/ Δ	
a Isolated by recrystallization (68%).							

phoshorus ylides were found to react with ethyl glyoxalate. Treatment of a chloroform solution of ylide 4 with 1 equiv of ethyl glyoxalate gave corresponding 1-azadienes 6 in moderate to good yields (Scheme 2, Table 2). Formation of 1-azadienes 6

Table 2. Polyfluorophosphorylated Derivatives 6 Obtained

entry	compd	R_{E}	Ar	reaction conditions	yield, $%$ ^{α}	
	6а	CF ₃	p -MeO-C ₆ H ₄	15 h/room temp	76	
2	6b	CF ₃	p -Me-C ₆ H ₄	12 h /room temp	65	
3	6с	CF ₃	$p\text{-NO}_2\text{-}C_6H_4$	$20 h$ /room temp	80	
4	6d	C_2F_5	$p\text{-NO}_2\text{-}C_6H_4$	24 h/reflux	68	
5	6e	CF ₃ H	$p\text{-}NO_2\text{-}C_6H_4$	15 h/room temp	76	
^a Isolated by flash chromatography.						

could be explained by a Wittig reaction between the ylide and the carbonyl group affording stereoselectively the corresponding E isomer of 1-azadiene and the corresponding phosphine oxide (Scheme 2).

Polyfluorophosphorylated 1-azadienes 6 were fully characterized by 1D and 2D NMR spectroscopy and MS spectrometry. One characteristic signal for azadiene 6a (R_F = CF_3 , Ar = p-MeOC₆H₄) in the ^IH NMR spectrum is the doublet at δ 6.96 ppm, with the coupling constant ${}^{3}J_{HP} = 22.7$ Hz, corresponding to the vinylic CH proton of the azadiene. The ¹³C NMR spectrum shows a characteristic doublet signal at δ 139.7 ppm with the coupling constant δ _{CP} = 7.6 Hz for the vinylic CH carbon and a doublet at δ 163.1 ppm with the coupling constant ${}^{3}J_{CP}$ = 26.0 Hz for the carboxyl carbon. Coupling

Scheme 2. Formation of Fluorinated Phosphorus Ylides 4 and 1-Azadienes 6

constants in this range are consistent with an E configuration of the vinylic double bond.

The potential conversion of functionalized azadienes 6 to fluorinated β -aminophosphonates was explored. Treatment of 6 with NaBH₄ at 0 °C afforded β -aminophosphonates 7/7' in good yields as syn/anti mixtures of diastereoisomers (Scheme 3, Table 3),

Scheme 3. Formation of Diastereomeric Mixture of Fluorinated β-Aminophosphonates by 1-Azadiene Reduction with N a $BH₄$

Table 3. Polyfluorophosphorylated Derivatives 7 Obtained

 a Isolated by flash chromatography. b Obtained as a diastereomeric mixture 60/40. ^cObtained as a diastereomeric mixture 95/5.

in different proportions for the syn and anti isomers, determined by NMR spectroscopy and mass spectrometry.

For example, the ³¹P NMR spectrum of β -aminophosphonates 7b/7′b ($R_F = CF_3$, $Ar = p$ -Me-C₆H₄) shows two singlets at δ 27.4 and 27.0 ppm, for the major and minor diastereomers (ratio 60/40), respectively, and in the 19F NMR spectrum two doublets appear at δ –75.6 ppm (³J_{FH} = 6.1 Hz) and at δ -72.4 ppm (3 J_{FH} = 7.6 Hz), for the major and minor diastereomers, respectively. The ¹H NMR spectrum shows as characteristic signals two methyl singlets at δ 2.15 ppm (major) and δ 2.14 ppm (minor) and two differentiated multiplets at 4.46− 4.56 ppm (major) and 4.26–4.38 ppm (minor) for the proton α to the CF_3 group. The ¹³C NMR spectrum shows two characteristic double quadruplets at δ 55.9 ppm with the two coupling constants $^2J_{CF} = 29.5$ Hz and $^2J_{CP} = 2.5$ Hz for the major isomer and at δ 56.2 ppm with the two coupling constants $^2J_{CF}$ = 30.5 Hz and $2J_{\text{CP}}$ = 4.3 Hz for the minor isomer, corresponding to carbons bonded to the CF_3 group. The anti or syn configuration assignment for β -aminophosphonates 7 and 7' was difficult to determine by NMR spectroscopy. Small differences in chemical shift, the overlap of signals, and complex splitting patterns due to the presence of nuclei such as phosphorus and fluorine in the structure precluded measurement of accurate coupling constants for each diastereoisomer.

Fortunately, in the case of β -aminophosphonate 7b (R_F = CF₃, $Ar = p$ -Me-C₆H₄), the isolation of the major *syn* diastereoisomer was possible in 46% yield by recrystallization from heptane (the minor diastereoisomer has significantly higher solubility), its structure being unequivocally determined by X-ray analysis (see the Supporting Information).

Formation of β -aminophosphonates 7 could be explained by red[uction of both the carb](#page-7-0)on−carbon and the iminic carbon− nitrogen double bonds to give both diastereoisomers of saturated β -aminophosphonates. The mechanism of this reaction could be explained as follows: first reduction of the carbon−carbon double bond by means of an initial attack of hydride at the most positive carbon of the heterodienic system to afford the corresponding enamine 8 (Scheme 3, vide supra), which is in tautomeric equilibrium with imine 9 and enamine 10, and subsequent reduction of the second double bond.

The relative energies of structures 8–10 (when $R_F = CF_3$, C_2F_5 , CF_2H) were calculated using Gaussian 09^{26} within the density functional theory (DFT) framework²⁷ at the B3LYP-(PCM)/6-31G* and M06-2X(PCM)//6-31G*//B[3L](#page-8-0)YP/6-31G* level using ethanol as solvent (for details [see](#page-8-0) the Supporting Information). Taking into account the results obtained, imines 9 are more stable than enamines 8 or 10 in all cases ($R_F = CF_3$, C_2F_5 , $CF₂H$; see the Supporting Information for details). Afterward, for the subsequent reduction of the iminic double bond of derivatives 9, a second hy[dride entrance to the im](#page-7-0)inic intermediate could be possible by a or b face approach (Scheme 4), affording the syn (7)

Scheme 4. Hydride Attack onto Imine 9

or anti (7′) diastereoisomer, respectively, of the corresponding β -aminophosphonate.

Hydride approaches the face containing the smaller groups, affording the corresponding syn β -aminophosphonate 7 as the major diastereoisomer. Therefore, experimental and theoretical results may explain that reduction of polyfluorophosphorylated 1-azadienes 6 with NaB H_4 in ethanol affords the corresponding syn β-aminophosphonates 7 as major diastereoisomers. It is noteworthy that these compounds show an interesting structure from a biological point of view, $2,6$ and no precedents for the synthesis of this type of polyfluoro β -aminophosphonates can be found in the literat[ure](#page-7-0).

β-Aminophosphonates 7 were efficiently converted to highly valuable γ -lactams^{17,18} by ring closure of these acyclic compounds. Base treatment of diastereomeric mixture of fluorinated β -aminophospho[nat](#page-7-0)[es](#page-8-0) $7/7'$ with NaH in THF afforded exclusively the cyclic trans-γ-lactam 11 as a single diastereoisomer (Scheme 5, Table 4).

The exclusive formation of trans-γ-lactam diastereoisomer [11](#page-3-0) may [be](#page-3-0) explained by deprotonation of the amine group with NaH in THF and intramolecular attack of the resulting nitrogen anion on the carbonyl group followed by the loss of ethanol to afford γ-lactams $11/11'$ as a mixture of *trans* and *cis* diastereoisomers. However, due to the basic reaction conditions, isomerization may occur in γ -lactams 11/11', and

Table 4. Polyfluorophosphorylated γ-Lactams 11 Obtained

only the most stable trans diastereomer of polyfluorophosphorylated-γ-lactam 11 (Scheme 5) is obtained. As far as we know, this strategy appears to represent the first synthesis of polyfluorophosphorylated γ-lactams.17,18

The structure of γ -lactams 11 was assigned on the basis of 1D and 2D NMR spectroscopy, inc[lud](#page-7-0)[in](#page-8-0)g HMQC and HMBC experiments, and mass spectrometric data. The ¹H NMR spectrum of compound 11c ($R_F = CF_3$, $Ar = p-NO_2-C_6H_4$) shows one double quadruplet at δ 4.90 ppm with the coupling constants ${}^{3}J_{\text{HF}} = 6.4$ and ${}^{3}J_{\text{HP}} = 18.4$ Hz corresponding to the proton 4-H bonded to the carbon with a CF_3 substituent (Figure 1); also the characteristic absence of carboxylic ethyl

Figure 1. Labeled and numbered carbons for γ -lactam 11c.

group signals is observed. Moreover, in the ³¹P NMR spectrum a singlet at δ 26.4 ppm and in the ¹⁹F NMR spectrum a doublet at δ -75.9 ppm with the coupling constant $4J_{\text{PF}}$ = 6.1 Hz are observed. The structure of compound 11c was also confirmed unambiguously by X-ray analysis (see the Supporting Information).

To have a better understanding of factors controlling the stereoselective formation of trans-γ-lactam 11[, the relative stabilitie](#page-7-0)s of cis-γ-lactam 11′ and trans-γ-lactam 11 were determined. Calculated free energy differences computed at the B3LYP(PCM)/ 6-31G* and M06-2X(PCM)//6-31G*//B3LYP/6-31G* level using tetrahydrofuran as solvent indicate that trans-γ-lactams 11a−e are about 3 kcal/mol more stable than cis-γ-lactams 11′a−e (Scheme 6 and the Supporting Information), and these

Scheme 6. Transformation of Compounds 11′ into 11 and Their Calculated Free Energy Differences (in kcal/mol) Computed at the B3LYP $(PCM)/6-31G^* + ZPVE$ Level Using Tetrahydrofuran as Solvent^a

ΔG $Ar - h$ $P(OEt)_2$ R_F 11		$Ar - p$ R_F 11'	$P(OEt)_2$
R_F	Ar	AG (kcal/mol)	
CF ₃	p-MeO-Ph	$-3.6(-3.0)$	
CF ₃	p-Me-Ph	-3.0 (-2.4)	
CF ₃	p -NO ₂ -Ph	$-3.5(-2.6)$	
C_2F_5	p -NO ₂ -Ph	$-3.9(-4.0)$	
CF ₂ H	p -NO ₂ -Ph	$-1.5(-2.4)$	

a Numbers in parentheses correspond to calculated free energy differences (in kcal/mol) computed at the M06-2X(PCM)/6- 31G*//B3LYP/6-31G* + ZPVE level using tetrahydrofuran as solvent.

results are in agreement with the isomerization of the γ -lactam under the basic reaction conditions.

■ CONCLUSION

In summary, we report the synthesis of functionalized polyfluorophosphorylated 1-azadienes by Wittig reactions from perfluorophosphorylated conjugated phosphoranes, obtained by the reaction of phosphazenes and fluorinated acetylenic phosphonates. Subsequent reduction of both carbon−carbon and carbon− nitrogen double bonds of 1-azadienes constitute a convenient synthetic route leading to novel fluoroalkyl β -aminophosphonates as a diastereomeric mixture, with the syn β -aminophosphonate being obtained as the major diastereoisomer. It was shown that base-mediated cyclocondensation of diastereomeric mixture of aminophosphonates leads exclusively to a new type of functionalized trans-γ-lactams. Fluoroalkyl β-aminophosphonates and polyfluorophosphorylated γ-lactams could be interesting, because these new phosphorus- and fluorine-containing compounds show promise for the preparation of novel, biologically active compounds useful in drug design.^{2,6,17,18} Theoretical calculations have been used to explain the diastereoselectivity of the 1-azadiene reduction mechanism, and co[mputa](#page-7-0)[tio](#page-8-0)nal studies also show a higher stability for the trans γ-lactams than for the cis isomers, in concert with the experimental observations.

EXPERIMENTAL SECTION

All reagents from commercial suppliers were used without further purification. All solvents were freshly distilled before use from appropriate drying agents. THF was distilled from sodium/benzophenone and used immediately. All other reagents were recrystallized or distilled when necessary. Reactions were performed under a dry nitrogen atmosphere. Analytical TLCs were performed with silica gel 60 F_{254} plates. Visualization was accomplished by UV light. Column chromatography was carried out using silica gel 60 (230−400 mesh ASTM). Melting points were determined with an electrothermal digital melting point apparatus without correction. NMR spectra were obtained on 300 MHz and 400 MHz spectrometers and recorded at 25 °C. Chemical shifts for ¹ ¹H NMR spectra are reported in ppm downfield from TMS, chemical shifts for ¹³C NMR spectra are recorded in ppm relative to internal chloroform (δ 77.2 ppm for ¹³C), chemical shifts for ¹⁹F NMR are reported in ppm downfield from fluorotrichloromethane $(CFCI₃)$, and chemical shifts for 31P NMR are reported in ppm from an aqueous solution of H_3PO_4 (33%). Coupling constants (J) are reported in hertz. The terms m, s, d, t, and q refer to multiplet, singlet, doublet, triplet, and quartet, respectively; br refers to a broad signal. 13 C NMR, 31 P NMR, and ¹⁹F NMR were broad-band decoupled from hydrogen nuclei. Infrared spectra (IR) were recorded with an infrared spectrometer; absorbance frequencies are given at maximum intensity in cm⁻¹. MS spectra were obtained on a chromatographic spectrometer, and HRMS spectra were

obtained on an instrument at 70 eV with a ionization source.
Perfluoroacetylene phosphonates²⁶ and azides²³ (*caution!* lowmolecular-weight carbon azides used in this study are potentially explosive; appropriate protection [mea](#page-8-0)sures shoul[d a](#page-8-0)lways be taken when handling these compounds) were prepared according to literature procedures.

General Procedure for the Preparation of Ylides. To a solution of azide (10 mmol) in CHCl₃ (25 mL) at 0 $^{\circ}$ C under nitrogen atmosphere was added dropwise a solution of phosphine (11 mmol). (Caution! Low-molecular-weight carbon azides used in this study are potentially explosive. Appropriate protection measures should always be taken when handling these compounds.) The mixture was stirred at room temperature until N_2 release had finished. Afterward a solution of the corresponding perfluoroacetylenephosphonate (10 mmol) in CHCl₃ was added dropwise and stirred at the corresponding temperature until the disappearance of starting materials was observed by $3^{1}P$ NMR spectra.

[1-Diethoxyphosphinyl-2-(4-tolylimino)-3,3,3-trifluoropropylidene] triphenylphosphorane (4g). The general procedure was followed using p-tolyl azide (1.33 g) and triphenylphosphine (2.62-g) stirred at 20 °C for 30 min followed by addition of trifluoromethylacetylenephosphonate 2a (2.31 g). (Caution! Low-molecular-weight carbon azides used in this study are potentially explosive. Appropriate protection measures should always be taken when handling these compounds.) The mixture was then stirred with refluxing chloroform for 21.5 h. Evaporation of the solvent gave a residue which was recrystallized in hexanes, affording 2.14 g of a yellowish solid (68%), mp 118–120 °C. ¹H NMR (300 MHz, CDCl₃): δ 1.08 (t, ³J_{HH} = 7.0 Hz, 6H, CH₃), 2.15 (s, 3H, CH₃), 3.76–3.83 (m, 4H, OCH₂), 5.67 (d, ³*I_{HH}* = 8.1 Hz, 2H, H_{arom}), 6.77 (d, ³*I_{HH}* = 8.1 Hz, 2H, H_{arom}), 7.43–7.55 (m, 10H, H_{arom}), 7.83–7.89 (m, 5H, H_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 16.2–16.4 (m, 2CH₃), 20.6 (CH₃), 60.7 (m, 2OCH₂), 118.2 (dq, ¹J_{CF} = 116.8 Hz, ³J_{CP} = 14.1 Hz, CF₃), 118.3 (HC_{arom}) , 118.5 (HC_{arom}), 123.0 (d, ³ J_{CP} = 17.1 Hz, C_{arom}), 126.2 (dd, ¹L – 25 Hz, C), 128.1 (HC), 128.2 (HC) $J_{\rm CP}$ = 91.7 Hz, $^{1}J_{\rm CP}$ = 2.5 Hz, C_{arom}), 128.1 (HC_{arom}), 128.2 (HC_{arom}), 128.3 (HC_{arom}), 128.4 (HC_{arom}), 128.6 (HC_{arom}), 129.1 (HC_{arom}), 130.8 (HC_{arom}) , 131.5 (HC_{arom}) , 131.6 (HC_{arom}) , 131.8 (HC_{arom}) , 131.8 (HC_{arom}) , 131.9 (HC_{arom}), 132.5 (d, ¹J_{CP} = 9.6 Hz, HC_{arom}), 133.8 (d, ¹L_{arom}), 147.8 (d, ¹L_{arom}), 153.4 (d, ²L $J_{\rm CP}$ = 9.6 Hz, HC_{arom}), 147.3 (C_{arom}), 146.6 (C_{arom}), 153.4 (dq, ²J_{CF} = 28.4 $\text{Hz, }^2I_{\text{CP}} = 9.2 \text{ Hz, } \text{C=N}$) ppm. ³¹P NMR (120 MHz, CDCl₃): δ 24.2 (d, $\frac{2I}{I} = 511 \text{ Hz}$) $\frac{2I}{I} = 511 \text{ Hz}$) ppm. ¹⁹E NMR (282 MHz $J_{\text{PP}} = 51.1 \text{ Hz}$), 25.7 (d, $^{2}J_{\text{PP}} = 51.1 \text{ Hz}$) ppm. ¹⁹F NMR (282 MHz, CDCl3): δ −59.3 ppm. MS (EI, 70 eV): m/z (%) 597 (10) [M]+ . HRMS

for $C_{32}H_{32}F_3NO_3P_2$ [M]⁺: calcd 597.1810, found 597.1815.
Hydrolysis of Ylides. Diethyl (3,3,3-Trifluoro-2-((4-methoxyphenyl)imino)propyl)phosphonate (5) and Diethyl (3,3,3-Trifluoro-2- ((4-methoxyphenyl)amino)prop-1-en-1-yl)phosphonate (5′). To a solution of 4-methoxyphenyl azide (10 mmol, 1.49 g) in CHCl₃ (25 mL) at 0 °C under a nitrogen atmosphere was added dropwise a solution of trimethylphosphine (11 mmol, 2.62 g). (Caution! Low-molecular-weight carbon azides used in this study are potentially explosive. Appropriate protection measures should always be taken when handling these compounds.) The mixture was then stirred in chloroform at room temperature for 30 min. Afterward a solution of trifluoromethylacetylenephosphonate $2a(10 \text{ mmol}, 2.31 \text{ g})$ in CHCl₃ was added dropwise and the mixture was stirred at room temperature for 30 min. To the resulting solution was added 1.5 mL of water, and the reaction mixture was stirred at room temperature for 1.5 h until consumption of starting ylide, as monitored by 31P NMR and 19F NMR spectroscopy. Evaporation of the solvent gave a residue which was purified by column chromatography, affording the corresponding mixture of imine 5 and enamine $5'$ as a colorless oil (70%), $R_f = 0.48$ (50/50, hexane/ethyl acetate). ¹H NMR (300 MHz, CDCl₃): δ 1.29–1.41 (m, 12H, CH₃), 3.14 (d, ²J_{HP} = 23.8 Hz, CH₂– P), 3.80 (s, 3H, OCH₃), 3.82 (s, 3H, OCH₃), 4.08−4.21 (m, 8H, OCH₂), 4.68 (d, ²J_{HP} = 6.0 Hz, =CH-P), 6.61 (d, ³J_{HH} = 7.1 Hz, 2H, H_{arom}), 6.79

 $(d, {}^{3}J_{HH} = 7.1 \text{ Hz}, 2H, H_{arom}$, 6.92 $(d, {}^{3}J_{HP} = 8.8 \text{ Hz}, 2H, H_{arom}$, 7.17 $(d, {}^{3}J_{HP} = 8.8 \text{ Hz}, 2H, H_{arom}$, 7.17 $(d, {}^{3}J_{HP} = 8.8 \text{ Hz}, 2H, H_{arom} = 8.8 \text{ Hz}$ ${}^{3}J_{\text{HP}}$ = 8.8 Hz, 2H, H_{arom}), 8.78 (s, NH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 15.8 (CH₃ imine), 16.1 (CH₃ enamine), 28.4 (d, ¹J_{CP} = 136.2 Hz, CH₂-P imine), 52.2 (d, ¹J_{CP} = 103.3 Hz, =C−P enamine), 55.3 (OCH₃ enamine), 55.4 (OCH₃ imine), 62.6 (d, ²J_{CP} = 6.1 Hz, OCH₂ enamine), 63.2 (d, ${}^{2}J_{CP} = 6.5$ Hz, OCH₂ imine), 113.9 (2HC_{arom} enamine), 114.3 (2HC_{arom} imine), 118.2 (q, $^{1}J_{CF}$ = 279.4 Hz, CF₃ enamine), 118.5 (q, $^{1}J_{CF}$ = 278.8 Hz, CF₃ imine), 120.9 (2HC_{arom} imine), 128.1 (2HC_{arom} enamine), 131.9 (d, $^{4}J_{CP} = 9.1$ Hz, C_{arom}-N enamine), 139.6 (C_{arom}–N imine), 139.7150.9 (m, C–CF₃ imine), 157.7 (C_{arom}–O imine), 158.1 (C_{arom}-O enamine), 160.5 (N-C= enamine) ppm. ³¹P NMR (120 MHz, CDCl₃): δ 19.6 (imine), 22.8 (enamine) ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –64.04 (enamine), -71.46 (imine) ppm. HRMS for $C_{14}H_{19}F_3NO_4P$ [M]⁺: calcd 353.1004, found 353.1009.

General Procedure for the Preparation of 1-Azadienes. To a solution of ylide (10 mmol) in CHCl₃ (25 mL) at 0 $^{\circ}$ C under a nitrogen atmosphere was added dropwise a solution of ethyl glyoxalate (10 mmol, 50% in toluene, 1.0 mL). The reaction mixture was stirred at the corresponding temperature and monitored by ³¹P NMR and ¹⁹F NMR spectroscopy until consumption of starting materials. Evaporation of the solvent gave a residue which was purified by column chromatography, affording the corresponding 1-azadienes.

(2E)-Ethyl 3-Diethoxyphosphinyl-5,5,5-trifluoro-4-(4-methoxyphenylimino)pent-2-enoate (6a). The general procedure was followed using ylide 4a and stirring at room temperature for 15 h. Evaporation of the solvent gave a residue which was purified by chromatography with hexane/ethyl acetate $(5/1)$ as eluent, affording 3.32 g of a orange oil (76%), $R_f = 0.43$ (50/50, hexane/ethyl acetate). H NMR (300 MHz, CDCl₃): δ 1.03 (t, ³J_{HH} = 7.2 Hz, 3H, CH₃), 1.21−1.42 (m, 6H, CH3), 3.74 (s, 3H, OCH3), 3.84−4.17 (m, 4H, OCH₂), 4.20–4.29 (m, 2H, OCH₂), 6.80 (d, ³J_{HH} = 8.8 Hz, 2H, H_{arom}), 6.93 (d, 3 J_{HH} = 8.8 Hz, 2H, H_{arom}), 6.96 (d, 3 J_{HP} = 22.7 Hz, 1H, $=$ CH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 13.7 (2CH₃), 15.7 $(2CH_3)$, 55.2 (OCH₃), 62.1 (OCH₂), 62.5 (d, ²J_{CP} = 6.2 Hz, OCH₂), 62.8 (d, $^2J_{CP}$ = 6.2 Hz, OCH₂), 113.8 (2HC_{arom}), 119.1 (q, $^1J_{CF}$ = 279.4 Hz, CF₃), 122.4 (2HC_{arom}), 138.7 (C_{arom}), 139.7 (d, ²J_{CP} = 7.6 Hz, HC=), 150.0 (dq, ² J_{CF} = 35.9 Hz, ² J_{CP} = 5.5 Hz, C-CF₃), 159.0 (C_{arom}) , 163.1 (d, 3 J_{CP} = 26.0 Hz, C=O) ppm. ³¹P NMR (120 MHz, CDCl₃): δ 9.6 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –68.9 ppm. HRMS for $C_{18}H_{23}F_3NO_6P [M]$ ⁺: calcd 437.1215, found 437.1214.

(2E)-Ethyl 3-Diethoxyphosphinyl-5,5,5-trifluoro-4-(4-tolylimino) pent-2-enoate (6b). The general procedure was followed using ylide 4b and stirring at room temperature for 15 h. Evaporation of the solvent gave a residue which was purified by chromatography with hexane/ethyl acetate (5/1) as eluent, affording 3.06 g of a yellowish oil (65%), $R_f = 0.43$ (50/50, hexane/ethyl acetate). ¹H NMR (300 MHz, CDCl₃): δ 1.05 (t, ³J_{HH} = 7.2 Hz, 3H, CH₃), 1.21–1.32 (m, 6H, CH₃), 2.27 (s, 3H, CH3), 3.92−4.10 (m, 4H, OCH2), 4.23−4.29 (m, 2H, OCH₂), 6.80 (d, ³J_{HH} = 6.5 Hz, 2H, H_{arom}), 6.90 (d, ³J_{HH} = 22.2 Hz, 1H, =CH), 7.07 (d, 3 J_{HH} = 6.5 Hz, 2H, H_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 13.7 (CH₃), 15.8 (CH₃), 15.9 (CH₃), 20.7 (CH₃), 62.1 (OCH₂), 63.0 (d, ²J_{CP} = 6.5 Hz, OCH₂), 63.2 (d, ²J_{CP} = 6.0 Hz, OCH₂), 118.4 (q, ¹J_{CF} = 279.5 Hz, CF₃), 120.0 (2HC_{arom}), 129.1 $(2HC_{arom})$, 135.8 (C_{arom}), 136.2 (d, ¹J_{CP} = 156.1 Hz, C−P), 139.6 (d, ³J_{CP} = 7.5 Hz, HC=), 144.3 (C_{arom}), 151.6 (dq, $^{2}J_{\text{CF}}$ = 36.6 Hz, $^{2}J_{\text{Cp}}$ = 5.5 Hz, C−CF₃), 163.2 (d, ³J_{CP} = 25.7 Hz, C=O) ppm. ³¹P NMR (120 MHz, CDCl₃): δ 9.4 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –69.1 ppm. HRMS for $C_{18}H_{23}F_3NO_5P$ [M]+: calcd 421.1266, found 421.1268.

(2E)-Ethyl 3-Diethoxyphosphinyl-5,5,5-trifluoro-4-(4-nitrophenylimino)pent-2-enoate (6c). The general procedure was followed using ylide 4c and stirring at room temperature for 20 h. Evaporation of the solvent gave a residue which was purified by chromatography with ether as eluent, affording 1.82 g of a yellowish oil (80%).

The general procedure was followed using ylide 4f and stirring at room temperature for 6 days. Evaporation of the solvent gave a residue which was purified by chromatography with ether as eluent, affording 0.68 g of a yellowish oil (30%), $R_f = 0.27$ (50/50, hexane/ethyl acetate). ¹H NMR (300 MHz, CDCl₃): δ 1.19−1.38 (m, 9H, CH₃), 4.02−4.14 (m, 4H, OCH₂), 4.31–4.38 (m, 2H, OCH₂), 6.80 (d, ³J_{HP} = 21.7 Hz, 1H, = CH), 7.03 (d, ${}^{3}J_{\text{HH}}$ = 8.9 Hz, 2H, H_{arom}), 8.22 (d, ${}^{3}J_{\text{HH}}$ = 8.8 Hz, 2H, H_{arom})

ppm. ¹³C NMR (75 MHz, CDCl₃): δ 13.9 (2CH₃), 16.1 (CH₃), 62.6 (OCH₂), 63.5 (d, ²J_{CP} = 17.1 Hz, OCH₂), 63.7 (d, ²J_{CP} = 17.1 Hz, OCH₂), 119.9 (2HC_{arom}), 124.7 (2HC_{arom}), 130.1 (q, ¹J_{CF} = 224.8 Hz, CF₃), 138.8 (d, ¹J_{CP} = 174.5 Hz, = C−P), 139.5 (d, ²J_{CP} = 7.1 Hz, HC=), 145.5 (C_{arom}), 152.7 (C_{arom}), 155.2 (dq, $^{2}J_{\text{CF}} = 37.4$ Hz, $^{2}J_{\text{CP}} =$ 8.9 Hz, C−CF₃), 163.5 (d, 3 J_{CP} = 25.2 Hz, C=O) ppm.³¹P NMR (120 MHz, CDCl₃): δ 9.2 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –69.7 ppm. HRMS for $C_{17}H_{20}F_3N_2O_7P$ [M]⁺: calcd 452.0960, found 452.0980.

(2E)-Ethyl 3-Diethoxyphosphinyl-3,3,4,4,4-pentafluoro-4-(4-nitrophenylimino)pent-2-enoate (6d). The general procedure was followed using ylide 4i and stirring in refluxing chloroform for 24 h. Evaporation of the solvent gave a residue which was purified by chromatography with hexane/ethyl acetate $(5/1)$ as eluent, affording 3.41 g of an orange oil (68%), $R_f = 0.72$ (50/50, hexane/ethyl acetate). ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$: δ 1.21−1.74 (m, 9H, CH₃), 3.82−4.21 (m, 4H, CH₂), 4.21–4.43 (m, 2H, CH₂), 6.82 (d, ³J_{HP} = 21.7 Hz, 1H, = CH), 7.07 (d, ${}^{3}J_{\text{HH}}$ = 8.9 Hz, 2H, H_{arom}), 8.21 (d, ${}^{3}J_{\text{HH}}$ = 8.8 Hz, 2H, H_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ13.9 (2CH₃), 16.1 (CH₃), 62.6 (OCH₂), 63.5 (d, ²J_{CP} = 17.1 Hz, OCH₂), 63.7 (d, ²J_{CP} = 17.1 Hz, OCH₂), 114.0−120.5 (m, C₂F₅), 119.6(2HC_{arom}), 125.6 (2HC_{arom}), 137.6(d, $^1J_{CP}$ = 174.7 Hz, =C-P), 139.6(d, $^2J_{CP}$ = 6.5 Hz, HC=), 145.3 (C_{arom}), 152.4 (C_{arom}), 154.0 (m, C–C₂F₅), 163.1(d, ³)_{CP} = 24.7 Hz, C=O) ppm. ³¹P NMR (120 MHz, CDCl₃): δ 8.2 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –80.8 (CF₃), –118.3 (CF₂) ppm. HRMS for $C_{18}H_{20}F_{5}N_2O_7P$ [M]⁺: calcd 502.0926, found 502.0928.

(2E)-Ethyl 3-Diethoxyphosphinyl-5,5-difluoro-4-(4-nitrophenylimino)pent-2-enoate (6e). The general procedure was followed using ylide 4j and stirring at room temperature for 15 h. Evaporation of the solvent gave a residue which was purified by chromatography with hexane/ ethyl acetate (5/1) as eluent, affording 1.65 g of a dark orange oil (76%), $R_f = 0.61$ (50/50, hexane/ethyl acetate). ¹H NMR (300 MHz, CDCl₃): $δ$ 1.19−1.38 (m, 9H, CH₃), 3.87−4.38 (m, 6H, CH₂), 6.32 (t, ³J_{HH} = 52.5 Hz, 1H, CHF₂), 6.78 (d, $^{3}J_{HP}$ = 24.7 Hz, 1H, = CH), 7.03 (d, $^{3}J_{HH}$ = 8.9 Hz, 2H, H_{arom}), 8.20 (d, $^{3}J_{\text{HH}}$ = 8.9 Hz, 2H, H_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 14.2 (CH₃), 16.3 (2CH₃), 62.6 (OCH₂), 63.7 (d, ²J_{CP} = 5.5 Hz, 2OCH₂), 111.9 (t, ¹J_{CF} = 247.3 Hz, CF₂H), 120.0 (2HC_{arom}), 124.6 (2HC_{arom}), 136.7 (C_{arom}), 138.2 (d, ¹J_{CP} = 26.2 Hz, = C-P), 145.2 $(d, {}^{2}J_{CP} = 7.1 \text{ Hz, CH}), 153.7 \text{ (C}_{arom}), 159.8−162.0 \text{ (m, C–CF}_2H), 163.8$ $(d, {}^{3}J_{CP} = 25.7 \text{ Hz}, \text{ C=O}) \text{ ppm}. {}^{31}P \text{ NMR}$ (120 MHz, CDCl₃): δ 8.9 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –120.2 (dq, ²J_{FF} = 306.7 Hz, ²J_{FH} = 27.6 Hz) ppm. HRMS for $C_{17}H_{21}F_2N_2O_7P$ [M]⁺: calcd 434.1054, found 434.1073.

General Procedure for the Preparation of β -Aminophosphonates. To a solution of the 1-azadiene (5 mmol) in ethanol (15 mL) cooled to 0 °C under a nitrogen atmosphere was added sodium borohydride portionwise. The reaction mixture was stirred at the corresponding temperature and monitored by 31P NMR and 19F NMR spectroscopy until consumption of starting materials. Then the reaction mixture was diluted with a water/HCl (2 M)/methylene chloride mixture $(1/1/3)$ $(3 \times 50 \text{ mL})$; the aqueous phase was extracted with methylene chloride and dried over $MgSO_4$, and the crude product was purified by column chromatography to afford the corresponding β -aminophosphonate.

(3S,4R),(3R,4S)- and (3S,4S),(3R,4R)-Ethyl 3-Diethoxyphosphinyl-5,5,5-trifluoro-4-(4-methoxyphenylamino)pentanoate (7a/7′a). The general procedure was followed using 1-azadiene 6a and 2 equiv of sodium borohydride (0.38 g, 10 mmol). The reaction mixture was stirred at room temperature for 2 h. Evaporation of the solvent gave a residue which was purified by chromatography with ether as eluent, affording 1.27 g of an orange oil (60%) as a 60/40 diastereomeric mixture, $R_f = 0.52$ (50/50, hexane/ethyl acetate). Data for the major isomer 7a are as follows. ¹H NMR (300 MHz, CDCl₃): δ 1.12−1.29 (m, 9H, CH₃), 2.59−2.93 (m, 3H, CH₂, P−CH), 3.66 (s, 3H, OCH₃), 3.91–4.52 (m, 8H, OCH₂, CH, NH), 6.67−6.73 (m, 4H, CHarom) ppm. 13C NMR (75 MHz, CDCl3): δ 14.0 (CH_3) , 16.1 (CH_3) , 16.2 (CH_3) , 31.9 (CH_2) , 33.0 $(d, {}^{1}J_{CP} = 143.7 \text{ Hz}$, CH−P), 55.6 (OCH₃), 55.7 (dq, ²J_{CF} = 30.9 Hz, ²J_{CP} = 4.0 Hz, CH−CF₃), 61.2 (OCH₂), 62.2–62.6 (2OCH₂), 114.5–116 (4HC_{arom}), 125.2 (dq, J_{CF} = 284.8 Hz, $^{3}J_{\text{CP}}$ = 15.6 Hz, CF₃), 139.9 (C_{arom}), 154.0 (C_{arom}), 171.4 $(d, {}^{3}J_{CP} = 15.3 \text{ Hz}, \text{C=O})$ ppm. ${}^{31}P$ NMR (120 MHz, CDCl₃): δ 27.4 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –72.6 (d, ⁴J_{FH} = 7.6 Hz) ppm.

HRMS for $C_{18}H_{27}F_3NO_6P$ [M]⁺: calcd 441.1532, found 441.1528. Data for the minor isomer 7'a are as follows. ¹H NMR (300 MHz, CDCl₃): δ 1.12−1.29 (m, 9H, CH3), 2.59−2.93 (m, 3H, CH2, P−CH), 3.69 (s, 3H, OCH₃), 3.91–4.52 (m, 7H, OCH_{2,} CH), 4.76 (d, 3 _{HH} = 10.3 Hz, 1H₁ NH), 6.57 (d, ³J_{HH} = 9.0 Hz, 2H, CH_{arom}), 6.67–6.73 (m, 2H, CH_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 14.1 (CH₃), 16.1 (CH₃), 16.3 $(CH₃)$, 30.5 (CH₂), 33.5 (d, ¹J_{CP} = 143.6 Hz, CH–P), 55.6 (OCH₃), 57.1 $(dq, {}^{2}J_{CF} = 30.3 \text{ Hz}, {}^{2}J_{CP} = 3.9 \text{ Hz}, \text{ CH–CF}_3$, 61.3 (OCH₂), 62.4–62.8 $(2OCH₂)$, 114.5−116 (4HC_{arom}), 125.8 (dq, ¹J_{CF} = 284.2 Hz, ³J_{CF} = 4.7 Hz, CF₃), 126.5 (2HC_{arom}), 140.5 (C_{arom}), 153.0 (C_{arom}), 171.6 (d, ³J_{CP} 12.8 Hz, C=O) ppm. ^{31}P NMR (120 MHz, CDCl₃): δ 27.0 ppm. ^{19}F NMR (282 MHz, CDCl₃): δ –72.5 (d, ³J_{FH} = 6.1 Hz) ppm. HRMS for $C_{18}H_{27}F_3NO_6P$ [M]⁺: calcd 441.1532, found 441.1528.

(3S,4R),(3R,4S)- and (3S,4S),(3R,4R)-Ethyl 3-Diethoxyphosphinyl-5,5,5-trifluoro-4-(4-tolylamino)pentanoate (7b/7′b). The general procedure was followed using the 1-azadiene 6b (5 mmol) and 2 equiv of sodium borohydride (0.38 g, 10 mmol). The reaction mixture was stirred at room temperature for 1 h. Evaporation of the solvent gave a residue which was purified by chromatography with hexane/ ethyl acetate (50/50) as eluent, affording 1.06 g of a yellow oil (50%) as a 60/40 diastereomeric mixture: $R_f = 0.54$ (50/50, hexane/ethyl acetate). Data for the major isomer 7b are as follows. ¹H NMR (300 MHz, CDCl₃): δ 1.10−1.26 (m, 9H, CH₃), 2.15 (s, 3H, CH₃), 2.55− 2.92 (m, 3H, CH−P, CH₂), 3.88−4.13 (m, 6H, OCH₂), 4.38 (d, β_{HH} = 10.7 Hz, 1H, NH), 4.46−4.56 (m, 1H, CH−CF₃), 6.62 (d, 3 J_{HH} = 8.5 Hz, 2H, H_{arom}), 6.88–6.92 (m, 2H, H_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 14.0 (CH₃), 16.0 (CH₃), 16.2 (CH₃), 20.2 (CH_3) , 31.8 (CH_2) , 33.4 $(d, {}^{1}J_{CP} = 143.4 \text{ Hz}$, CH-P), 55.9 $(dq, {}^{2}J_{CF} =$ 29.5 Hz, ² J_{CP} = 2.5 Hz, CH–CF₃), 61.0 (d, ² J_{CP} = 3.8 Hz, OCH₂), 62.4 $(d, {}^{2}J_{CP} = 6.9 \text{ Hz}, \text{ OCH}_2), 114.0 \text{ (2CH}_{arom}), 125.5 \text{ (dq, } {}^{1}J_{CF} = 284.8$ Hz, ${}^{3}J_{CP}$ = 15.5 Hz, CF₃), 128.3 (C_{arom}), 129.6 (2CH_{arom}), 143.4 (C_{arom}) , 171.1 (d, ${}^{3}C_{\text{P}} = 15.3$ Hz, C=O) ppm. ³¹P NMR (120 MHz, CDCl₃): δ 27.4 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –75.6 (d, ${}^{3}L_{12} = 6.1$ Hz) ppm. HRMS for C₁.H₁-E-NO₁P [M]⁺; calcd 425.1579 $J_{\text{FH}} = 6.1 \text{ Hz}$) ppm. HRMS for $C_{18}H_{27}F_3NO_5P \text{ [M]}^+$: calcd 425.1579, found 425.1590. Data for the minor isomer $7'$ b are as follows. 1 H NMR (300 MHz, CDCl₃): δ 1.10–1.26 (m, 9H, CH₃), 2.14 (s, 3H, CH₃), 2.55−2.92 (m, 3H, CH−P, CH₂), 3.88−4.13 (m, 6H, OCH₂), 4.26−4.38 (m, 1H, CH−CF₃), 4.90 (d, ³J_{HH} = 10.4 Hz, 1H, NH), 6.50 (d, ${}^{3}J_{\text{HH}}$ = 8.4 Hz, 2H, H_{arom}), 7.00–7.17 (m, 2H, H_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 14.1 (CH₃), 16.1 (2CH₃), 20.1 (CH₃), 29.9 (CH₂), 33.3 (d, ¹J_{CP} = 143.5 Hz, CH-P), 56.2 (dq, ²J_{CF} = 30.5 Hz , 2 J_{CP} = 4.3 Hz, CH–CF₃), 61.2 (d, 2 J_{CP} = 6.3 Hz, OCH₂), 62.5 $(d, {}^{2}J_{CP} = 6.9 \text{ Hz}, \text{ OCH}_2)$, 113.3 (2CH_{arom}), 125.1 (dq, ${}^{1}J_{CF} = 284.3$ Hz, ${}^{3}J_{CP}$ = 4.3 Hz, CF₃), 127.8 (C_{arom}), 129.6 (2CH_{arom}), 143.9 (C_{arom}) , 170.9 (d, ${}^{3}C_{\text{P}} = 12.5 \text{ Hz}$, C=O) ppm. ³¹P NMR (120 MHz, CDCl₃): δ 27.0 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ -72.4 (d, $J_{\text{FH}} = 7.6 \text{ Hz}$) ppm. HRMS for $C_{18}H_{27}F_3NO_5P [M]^{+}$: calcd 425.1579, found 425.1590.

(3S,4R),(3R,4S)- and (3S,4S),(3R,4R)-Ethyl 3-Diethoxyphosphinyl-5,5,5-trifluoro-4-(4-nitrophenylamino)pentanoate (7c/7′c). The general procedure was followed using the 1-azadiene 6c (5 mmol) and 1 equiv of sodium borohydride (0.19 g, 5 mmol). The reaction mixture was stirred at room temperature for 1 h. Evaporation of the solvent gave a residue which was purified by chromatography with ether as eluent, affording 1.87 g of an orange oil (82%) as a 60/40 diastereomeric mixture: $R_f = 0.76$ (50/50, hexane/ethyl acetate). Data for the major isomer 7c are as follows. ¹H NMR (300 MHz, CDCl₃): δ 1.21–1.24 (m, 9H, CH₃), 2.57–3.02 (m, 3H, CH₂, P–CH), 4.04–4.26 (m, 6H, OCH₂), $4.53-4.85$ (m, 1H, CH), 6.13 (d, $3I_{\text{HH}} = 10.1 \text{ Hz}$, 1H, NH), 6.77 (d, $3I_{\text{H}} = 9.7 \text{ Hz}$, $2H_{\text{H}} = 1.809-8.14 \text{ (m)}$ and $H = 1 \text{ mm}$, 13 C NMR (75) ${}^{3}J_{\text{HH}}$ = 9.2 Hz, 2H, H_{arom}), 8.09–8.14 (m, 2H, H_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 14.3 (CH₃), 16.5 (CH₃), 16.6 (CH₃), 30.5 (CH₂), 33.5 $(d, {}^{1}J_{CP} = 143.6 \text{ Hz}, \text{HC-P})$, 53.4 $(dq, {}^{2}J_{CF} = 30.5 \text{ Hz}, {}^{2}J_{CP} = 3.3 \text{ Hz}, \text{HC}-$ CF₃), 61.9 (OCH₂), 61.2−62.5 (m, 2OCH₂), 112.6 (2HC_{arom}), 124.5 $(dq, {}^{1}J_{CF} = 284.9 \text{ Hz}, {}^{3}J_{CP} = 13.1 \text{ Hz}, \text{ CF}_3)$, 126.5 (2HC_{arom}), 139.7 (C_{arom}) , 151.6 (C_{arom}) , 171.4 $(d, {}^{3}J_{\text{CP}} = 15.6 \text{ Hz}, C=O)$ ppm. ${}^{31}P$ NMR (120 MHz, CDCl₃): δ 27.0 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –72.5 (d, $^{4}J_{\text{FH}}$ = 7.6 Hz) ppm. HRMS for $C_{17}H_{21}F_{2}N_{2}O_{7}P$ [M]⁺: calcd 434.1054, found 434.1061. Data for the minor isomer ⁷′^c are as follows. ¹ ¹H NMR (300 MHz, CDCl₃): δ 1.21–1.24 (m, 9H, CH₃), 2.57–3.02 (m, 3H, CH₂, CH), 4.04−4.26 (m, 6H, OCH₂), 4.53−4.85 (m, 1H, CH),

6.22 (d, ${}^{3}J_{\text{HH}}$ = 10.2 Hz, 1H, NH), 6.67 (d, ${}^{3}J_{\text{HH}}$ = 9.2 Hz, 2H, H_{arom}), 8.09−8.14 (m, 2H, H_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 14.3 (CH_3) , 16.3 (CH₃), 16.4 (CH₃), 31.9 (CH₂), 33.0 (d, ¹J_{CP} = 143.7 Hz, HC−P), 55.0 (dq, $^2J_{CF}$ = 31.6 Hz, $^2J_{CP}$ = 4.3 Hz, HC−CF₃), 61.8 (OCH₂), 61.2–62.5 (m, 2OCH₂), 112.2 (2HC_{arom}), 124.5 (dq, ¹J_{CF} = 288.5 Hz, ${}^{3}J_{CP} = 4.3$ Hz, CF₃), 126.5 (2HC_{arom}), 139.7 (C_{arom}), 152.0 (C_{arom}) , 171.6 (d, ${}^{3}J_{\text{CP}} = 13.2 \text{ Hz}$, C=O) ppm. ${}^{31}\text{P}$ NMR (120 MHz, CDCl₃): δ 26.9 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –72.1 (d, ⁴J_{FH} = 7.6 Hz) ppm. HRMS for $C_{17}H_{21}F_2N_2O_7P$ [M]⁺: calcd 434.1054, found 434.1063.

(3S,4R),(3R,4S)- and (3S,4S),(3R,4R)-Ethyl 3-Diethoxyphosphinyl-5,5,6,6,6-pentafluoro-4-(4-nitrophenylamino)hexanoate (7d/7′d). The general procedure was followed using the 1-azadiene 6d (5 mmol) and 2 equiv of sodium borohydride (0.38 g, 10 mmol). The reaction mixture was stirred at room temperature for 1 h. Evaporation of the solvent gave a residue which was purified by chromatography with ether as eluent, affording 1.52 g of a yellow oil (60%) as a $95/5$ diastereomeric mixture: $R_f = 0.52$ (50/50, hexane/ ethyl acetate). Data for the major isomer 7**d** are as follows. $^1\mathrm{H}$ NMR (300 MHz, CDCl₃): δ 1.20−1.37 (m, 9H, CH₃), 2.67−2.86 (m, 3H, CH2, CH), 3.92−4.27 (m, 6H, OCH2), 4.86−5.01 (m, 1H, CH), 5.71 $(d, {}^{3}J_{\text{HH}} = 11.1 \text{ Hz}, 1H, \text{NH}), 6.74 (d, {}^{3}J_{\text{HH}} = 9.2 \text{ Hz}, 2H, H_{\text{arom}}), 8.07$ $(d, {}^{3}J_{HH} = 9.2 \text{ Hz}, 2H, H_{arom}) \text{ ppm}. {}^{13}C \text{ NMR} (75 \text{ MHz}, \text{CDCl}_{3}): \delta$ 14.1 (CH₃), 16.2 (CH₃), 16.3 (CH₃), 30.3 (CH₂), 33.8 (d₁¹J_{CP} = 143.3 Hz, HC−P), 50.4–50.9 (m, HC−C₂F₅), 61.6 (OCH₂), 62.1– 62.8 (m, 2OCH₂), 112.4 (HC_{arom}), 121.1–126.3 (m, C₂F₅), 126.2 (HC_{arom}) , 139.7 (C_{arom}), 150.7 (C_{arom}), 171.4 (d, ³J_{CP} = 13.6 Hz, C= O) ppm. ^{31}P NMR (120 MHz, CDCl₃): δ 26.3 ppm. ^{19}F NMR (282) MHz, CDCl₃): δ -82.4 (CF₃), -116.4 (d, F_a, ²J_{FF} = 274.4 Hz), -126.3 (dd, F_b, 2 *J_{FF}* = 274.4 Hz, 3 *J_{FH}* = 24.0 Hz) ppm. HRMS for $C_{18}H_{24}F_5N_2O_7P$ [M]⁺: calcd 506.1239, found 506.1241. Data for minor isomer 7'd are as follows. ³¹P NMR (120 MHz, CDCl₃): δ 26.1 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –82.7 (CF₃), –118.0 (d, F_a, 2_{JFF} = 273.8 Hz), –124.2 (dd, F_b, ²J_{FF} = 273.8 Hz, ³J_{FH} = 22.0 Hz) ppm. HRMS for $C_{18}H_{24}F_5N_2O_7P$ [M]⁺: calcd 506.1239, found 506.1241.

(3S,4R),(3R,4S)- and (3S,4S),(3R,4R)-Ethyl 3-Diethoxyphosphinyl-5,5-difluoro-4-(4-nitrophenylamino)pentanoate (7e/7'e). The general procedure was followed using the 1-azadiene 6e (5 mmol) and 2 equiv of sodium borohydride (0.38 g, 10 mmol). The reaction mixture was stirred at room temperature for 2 h. Evaporation of the solvent gave a residue which was purified by chromatography with ether as eluent, affording 0.77 g of a yellow oil (35%) as a 95/5 diastereomeric mixture,: $R_f = 0.24$ (50/50, hexane/ethyl acetate). Data for major isomer 7e are as follows. ¹H NMR (300 MHz, CDCl₃): δ 1.17–1.57 (m, 9H, CH3), 2.45−2.98 (m, 3H, CH2, CH), 4.10−4.20 (m, 6H, OCH₂), 4.20–4.39 (m, 1H, CH), 5.81 (d, ³J_{HH} = 10.0 Hz, 1H, NH), 6.05 (dt, $^{2}J_{\text{HF}} = 57.7$ Hz, $^{3}J_{\text{HH}} = 6.1$ Hz, 1H, CF₂H), 6.45 (d, $^{3}J_{\text{HH}} =$ 10.3 Hz, 2H, CH_{arom}), 8.08 (d, ³J_{HH} = 10.3 Hz, 2H, CH_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 14.7 (CH₃), 16.4–16.2 (m, 2CH₃), 31.0 $(CH₂)$, 32.8 (d, ¹J_{CP} = 136.1 Hz, HC−P), 54.7–55.4 (m, HC−CF₂H), 61.5 (OCH₂), 62.9–63.2 (m, 2OCH₂), 111.9 (HC_{arom}), 115.4 (t, J_{CF} = 277.0 Hz, CF₂H), 126.2(HC_{arom}), 139.0 (C_{arom}), 152.4(C_{arom}), 170.8 (d, ${}^{3}I_{CP}$ = 16.6 Hz, C=O) ppm. ${}^{31}P$ NMR (120 MHz, CDCl₃): δ 27.5 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –121.0 (ddd, F_a, ²J_{FF} = 285.0 Hz, ²J_{FH} = 55.6 Hz, ³J_{FH} = 6.1 Hz), -124.1 (ddd, F_b, ²J_{FF} = 285.0 $\text{Hz}_{12}{}^{2}J_{\text{FH}} = 56.4 \text{ Hz}_{12}{}^{3}J_{\text{FH}} = 12.2 \text{ Hz}$ ppm. HRMS for $\text{C}_{17}\text{H}_{25}\text{F}_{2}\text{N}_{2}\text{O}_{7}\text{P}_{2}$ [M]+ : calcd 438.1367, found 438.1384. Data for minor isomer 7′e are as follows. 31P NMR (120 MHz, CDCl3): δ 27.4 ppm. 19F NMR (282 MHz, CDCl₃): δ –121.2 (dd, F_a, ²J_{FF} = 284.5 Hz, ³J_{FH} = 6.2 Hz), –124.4 (ddd, $F_{\rm b}$, $^2J_{\rm FF}$ = 284.5 Hz, $^2J_{\rm FH}$ = 56.4 Hz, $^3J_{\rm FH}$ = 12.9 Hz) ppm. HRMS for $C_{17}H_{25}F_{2}N_{2}O_{7}P$ [M]⁺: calcd 438.1367, found 438.1384.

General Procedure for the Preparation of γ -Lactams. To a sodium hydride suspension (0.04 g, 1.5 mmol) in THF (5 mL) cooled to 0 °C under a nitrogen atmosphere was added dropwise a solution of the β -aminophosphonate (1 mmol) in THF. The reaction mixture was stirred at the corresponding temperature and monitored by ${}^{31}P$ NMR and ${}^{19}F$ NMR spectroscopy until consumption of the starting β -aminophosphonate. Evaporation of the solvent gave a residue which was purified by column chromatography, affording the corresponding γ-lactam.

4-Diethoxyphosphinyl-5-trifluoromethyl-1-(4-methoxyphenyl) pyrrolidin-2-one (11a). The general procedure was followed using a diastereomeric mixture of β -aminophosphonates 7a/7'a (0.44 g). The reaction mixture was stirred at room temperature for 15 h. Evaporation of the solvent gave a residue which was purified by chromatography with hexane/ethyl acetate (1/10) as eluent, affording 0.36 g of an orange oil (90%): $R_f = 0.24$ (50/50, hexane/ethyl acetate). ¹H NMR (300 MHz, CDCl₃): δ 1.25−1.35 (m, 6H, CH₃), 2.66−3.08 (m, 3H, CH, CH₂), 3.56 (s, 3H, OMe), 4.11−4.22 (m, 4H, OCH₂), 4.55−4.65 $(m, 1H, CH-CF₃), 6.87 (d, ³)_{HH} = 8.9 Hz, 2H, H_{arom}), 7.16 (d, ³)_{HH} =$ 8.9 Hz, 2H, H_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 16.4 (CH₃), 16.4 (CH₃), 29.4 (d, ¹J_{CP} = 107 Hz, CH), 30.6 (d, ²J_{CP} = 4.7 Hz, CH₂), 55.4 (OCH₃), 61.0 (q, ²J_{CF} = 28.8 Hz, HC–CF₃), 63.3 (d, ²J_{CF} = 6.8 Hz, 2OCH₂), 114.6 (2HC_{arom}), 124.7 (dq, ¹J_{CF} = 268.3 Hz, ³J_{CF} = 15.7 Hz, CF₃), 127.1 (2HC_{arom}), 129.5 (C_{arom}), 158.9 172.6 (C=O) ppm. ³¹P NMR (120 MHz, CDCl₃): δ 26.5 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –76.0 (d, ⁴J_{FH} = 6.1 Hz) ppm. HRMS for $C_{16}H_{21}F_3NO_5P$ [M]⁺: calcd 395.1126, found 395.1109.

4-Diethoxyphosphinyl-5-trifluoromethyl-1-(4-tolyl)pyrrolidin-2 one (11b). The general procedure was followed using a diastereomeric mixture of β -aminophosphonates 7b/7'b (0.46 g). The reaction mixture was stirred at room temperature for 3 h. Evaporation of the solvent gave a residue which was purified by chromatography with hexane/ethyl acetate (10/1) as eluent, affording 0.37 g of an orange oil (80%) : $R_f = 0.29$ (50/50, hexane/ethyl acetate). ¹H NMR (300 MHz, CDCl₃): δ 1.20−1.24 (m, 6H, CH₃), 2.29 (s, 3H, CH₃), 2.70−3.10 (m, 3H, CH, CH₂), 4.11−4.18 (m, 4H, OCH₂), 4.87−4.93 (m, 1H, CH−CF₃), 7.12 (d, ³J_{HH} = 8.7 Hz, 2H, HC_{arom}), 7.16 (d, ³J_{HH} = 8.7 Hz, 2H, HC_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 16.4 (CH₃), 16.4 (CH₃), 29.2 (d, ¹J_{CP} = 150.6 Hz, CH), 31.0 (d, ²J_{CP} = 4.7 Hz, CH₂), 61.0 (q, ² J_{CF} = 34.2 Hz, C–CF₃), 63.3 (d, ² J_{CP} = 6.6 Hz, 2OCH₂), 124.1 (2HC_{arom}), 124.5 (dq, ¹J_{CF} = 284.3 Hz, ³J_{CP} = 16.0 Hz, CF_3), 124.8 (2HC_{arom}), 142.5 (C_{arom}), 145.7 (C_{arom}), 172.0 (C=O) ppm. 31P NMR (120 MHz, CDCl3): δ 26.4 ppm. 19F NMR (282 MHz, CDCl₃): δ –76.0 (d, ⁴J_{FH} = 6.1 Hz) ppm. MS (EI, 70 eV): m/z (%) 379 (3) [M⁺]. HRMS for $C_{16}H_{21}F_3NO_4P$ [M]⁺: calcd 379.1171, found 379.1160.

4-Diethoxyphosphinyl-5-trifluoromethyl-1-(4-nitrophenyl) pyrrolidin-2-one (11c). The general procedure was followed using a diastereomeric mixture of β -aminophosphonates 7c/7'c (0.46 g). The reaction mixture was stirred at room temperature for 24 h. Evaporation of the solvent gave a residue which was purified by chromatography with hexane/ethyl acetate $(10/1)$ as eluent, affording 0.36 g of a yellow solid (90%): mp 115−117 °C (methylene chloride/hexane). ¹ H NMR (300 MHz, CDCl₃): δ 1.24 (m, 6H, CH₃), 2.70–3.20 (m, 3H, CH, CH₂), 4.11−4.18 (m, 4H, OCH₂), 4.90 (dq, 1H, ${}^{3}J_{HF}$ = 6.4 Hz, ${}^{3}J_{HF}$ = 18.4 Hz, CH−CF₃), 7.55 (d, ³J_{HH} = 8.7 Hz, 2H, HC_{arom}), 8.23 (d, ³J_{HH} = 8.8 Hz, 2H, HC_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 16.4 (CH_3) , 16.4 (CH_3) , 29.2 $(d, {}^{1}J_{CP} = 150.6 \text{ Hz}$, CH), 31.0 $(d, {}^{2}J_{CP} = 4.7 \text{ s}$ Hz, CH₂), 61.0 (q, ²J_{CF} = 34.2 Hz, C–CF₃), 63.3 (d, ²J_{CP} = 6.6 Hz, 2OCH₂), 124.1 (2HC_{arom}), 124.5 (dq, ¹J_{CF} = 284.3 Hz, ³J_{CP} = 16.0 Hz, CF_3), 124.8 (2HC_{arom}), 142.5 (C_{arom}), 145.7 (C_{arom}), 172.0 (C=O) ppm. ³¹P NMR (120 MHz, CDCl₃): δ 26.4 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –75.9 (d, ⁴J_{FH} = 6.1 Hz) ppm. HRMS for C₁₅H₁₈F₃N₂O₆P [M]⁺: calcd 410.0855, found 410.0859.

4-Diethoxyphosphinyl-5-perfluoroethyl-1-(4-nitrophenyl) pyrrolidin-2-one (11d). The general procedure was followed using a diastereomeric mixture of β -aminophosphonates 7d/7'd (0.46 g). The reaction mixture was stirred at room temperature for 36 h. Evaporation of the solvent gave a residue which was purified by chromatography with hexane/ethyl acetate (10/1) as eluent, affording 0.37 g of an orange oil (90%): $R_f = 0.36$ (50/50, hexane/ethyl acetate). ¹H NMR (300 MHz, CDCl3): δ 1.22−1.34 (m, 6H, CH3), 2.74−3.21 (m, 3H, CH, CH₂), 4.12−4.25 (m, 4H, CH₂), 4.97−5.17 (m, 1H, CH−CF₂), 7.60 (d, ${}^{3}J_{\text{HH}} = 9.0$ Hz, 2H, HC_{arom}), 8.29(d, ${}^{3}J_{\text{HH}} = 9.0$ Hz, 2H, HC_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 16.2 (CH₃), 16.4 (CH_3) , 29.2 (d, ¹J_{CP} = 143.9 Hz, CH), 31.0 (d, ²J_{CP} = 6.8 Hz, CH₂), 59.5 (m, HC−CF₂), 63.3 (d, ²J_{CP} = 2.0 Hz, 2OCH₂), 111.2−116.7 (m, CF₃), 118.3 (qt, ${}^{1}J_{CF}$ = 305.9 Hz, ${}^{2}J_{CF}$ = 35.1 Hz, CF₂), 124.1 (2HC_{arom}), 124.6 (2HC_{arom}), 143.1 (C_{arom}), 145.6 (C_{arom}), 172.0 (C=O) ppm. ^{31}P NMR (120 MHz, CDCl₃): δ 25.6 ppm. ¹⁹F NMR (282 MHz, CDCl₃): δ –81.9 (CF₃), –123.4 (d, ²J_{FF} = 280.0 Hz, CF₃) ppm. HRMS for $C_{16}H_{18}F_5N_2O_6P$ [M]⁺: calcd 460.0834, found 460.0823.

4-Diethoxyphosphinyl-5-difluoromethyl-1-(4-nitrophenyl) pyrrolidin-2-one (11e). The general procedure was followed using a diastereomeric mixture of β -aminophosphonates 7e/7'e (0.44 g). The reaction mixture was stirred at room temperature for 5 h. Evaporation of the solvent gave a residue which was purified by chromatography with hexane/ethyl acetate $(10/1)$ as eluent, affording 0.27 g of an orange oil (70%): $R_f = 0.39$ (50/50, hexane/ethyl acetate). ¹H NMR (300 MHz, CDCl₃): δ 1.25−1.74 (m, 6H, CH₃), 2.66−3.12 (m, 3H, CH,CH2), 4.12−4.19 (m, 4H, 2OCH2), 4.78−4.98 (m, 1H, CH- CF_2H), 5.97 (t, ²J_{HF} = 54.5 Hz, 1H, CF₂H), 7.13 (d, ³J_{HH} = 8.9 Hz, 2H, HC_{arom}), 7.21(d, ${}^{3}\text{J}_{\text{HH}}$ = 8.9 Hz, 2H, HC_{arom}) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 16.4 (CH₃), 16.4 (CH₃), 29.9 (d, ²J_{CP} = 5.5 Hz, CH₂), 31.3 (d, ¹J_{CP} = 160.8 Hz, CH), 60.8–61.0 (m, HC–CF₂H), 63.3 (2OCH₂), 100.8−116.1 (m, CF₂H), 123.3 (HC_{arom}), 125.0 (HC_{arom}) , 142.5 (C_{arom}), 145.8 (C_{arom}), 172.3 (C=O) ppm. ³¹P NMR (120 MHz, CDCl₃): δ 26.8 ppm. ¹⁹F NMR (282 MHz, CDCl₃) δ -129.2 (dd, $^{2}J_{\text{FH}}$ = 7.6 Hz, $^{2}J_{\text{FH}}$ = 55.0 Hz) ppm. HRMS for $C_{15}H_{19}F_{2}N_{2}O_{6}P$ [M]⁺: calcd 392.0961, found 392.0949.

■ ASSOCIATED CONTENT

S Supporting Information

Figures, tables, and CIF files giving ¹H NMR and ¹³C NMR spectra of compounds 4g, 5/5′, 6a−e, 7/7′a−e, 7b, and 11a−e, 2D spectra of compounds 6a and 11c, ORTEP and X-ray crystallographic data of compounds 7b and 11c, and Cartesian coordinates, harmonic analysis data, and energies for all the stationary points discussed in the computational studies. This material is available free of charge via the Internet at http:// pubs.acs.org.

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■ ACKNOWLEDGMENTS

Financial support from the Dirección General de Investigación del Ministerio de Ciencia e Innovación (CTQ2009-12156) and by Gobierno Vasco and Universidad del País Vasco (GV, IT 422-10; UPV, UFI-QOSYC 11/12) is gratefully acknowledged. UPV/EHU-SGIker technical support (MICINN, GV/EJ, European Social Fund) is also gratefully acknowledged.

■ REFERENCES

(1) For reviews see: (a) Mucha, A.; Kafarski, P.; Berlicki, L. J. Med. Chem. 2011, 54, 5955−5980. (b) Palacios, F.; de los Santos, J. M.; Vicario, J.; Alonso, C., The Chemistry of Hydroxylamines, Oximes and Hydroxamic Acids - Patai's Chemistry of Functional Groups; Wiley: Chichester, U.K., 2011; Vol. 2, pp 351–439. (c) Ordóñez, M.; Rojas-Cabrera, H.; Cativiela, C. Tetrahedron 2009, 65, 17−49. (d) Aminophosphonic and Amino-phosphinic Acids. Chemistry and Biological Activity; Kukhar, V. P., Hudson, H. R., Eds.; Wiley: Chichester, U.K., 2000.

(2) For reviews see: (a) Van der Jeught, S.; Stevens, Ch. V. Chem. Rev. 2009, 109, 2672−2702. (b) Palacios, F.; Alonso, C.; de los Santos, J. M. Chem. Rev. 2005, 105, 899−931.

(3) (a) Metcaft, W. W.; van der Donk, W. A. Annu. Rev. Biochem. 2009, 78, 65−94. (b) Fields, S. C. Tetrahedron 1999, 55, 12237− 12273. (c) Horiguchi, M.; Kandatsu, M. Nature 1959, 184, 901−902. (d) Hammerschmidt, F. Liebigs Ann. Chem. 1988, 531−535. (e) Hammerschmidt, F.; Vollenkle, H. Liebigs Ann. Chem. 1989, 577−583. (f) Stalikas, C. D.; Konidani, C. N. J. Chromatogr. A 2001,

901, 1−19. (g) Cabeza, A.; Ouyang, X.; Sharma, C. V. K.; Aranda, M. A. G.; Bruque, S.; Clearfield, A. Inorg. Chem. 2002, 41, 2325−2333. (h) Caseiola, M.; Castantino, W.; Peraio, A.; Rega, T. Solid State Ionics 1995, 77, 229−233.

(4) (a) Ojima, I. In Fluorine in Medicinal Chemistry and Chemical Biology; Wiley-Blackwell: Chichester, U.K., 2009. (b) Begué, J. P.; Bonnet-Delpon, D. In Bioorganic and Medicinal Chemistry of Fluorine; Wiley: Chichester, U.K., 2008. (c) Kirsch, P. In Modern Fluoroorganic Chemistry. Synthesis, Reactivity, Application; Wiley-VCH: Weinheim, Germany, 2004.

(5) As many as 25−30% of agrochemicals and 20% of pharmaceuticals on the market are estimated to contain fluorine.

(6) For reviews see: (a) Vulpetti, A.; Dalvit, C. Drug Discovery Today 2012, 17, 890–897. (b) Müller, K.; Böhm, H.-J. Chem. Biol. 2009, 16, 1130−1131. (c) Filler, R.; Saha, R. Fut. Med. Chem. 2009, 1, 777−791. (d) Hagmann, W. K. J. Med. Chem. 2008, 51, 4359−4369. (e) Purser, S.; Moore, P. R.; Swallow, S.; Gouverneur, V. Chem. Soc. Rev. 2008, 37, 320−330. (f) Kirk, K. L. Org. Process Res. Dev. 2008, 12, 305−321. (g) Kirk, K. L. J. Fluorine Chem. 2006, 127, 1013−1029. (h) Dolbier, W. R. J. Fluorine Chem. **2006**, 126, 157−163. (i) Böhm, H.-J.; Banner, D.; Bendels, S.; Kansy, M.; Kuhn, B.; Müller, K.; Obst-Sander, U.; Stahl, M. l. ChemBioChem 2004, 5, 637−643. (j) Strunecka, A.; Patocka, J.; Connett, P. J. Appl. Biomed. 2004, 2, 141−150.

(7) (a) Modern Crop Protection Compounds, 2nd ed.; Kraemer, W., Schirmer, U., Jeschke, P., Witschel, M., Eds.; Wiley: Chichester, U.K., 2011; Vols. 1−3. (b) Jeschke, P. Pest Manage. Sci. 2010, 66, 10−27. (c) Theodoridis, G. Adv. Fluorine Sci. 2006, 2, 121−175. (d) Thayer, A. M. Chem. Eng. News 2006, 84, 15−17. (e) Jeschke, P. ChemBioChem 2004, 5, 570−589.

(8) Pagliaro, M.; Ciriminna, R. J. Mater. Chem. 2005, 15, 4981−4991. (9) Müller, K.; Faeh, C.; Diederich, F. Science 2007, 317, 1881−1886. (10) (a) Prabhakaran, J.; Underwood, M. D.; Parsey, R. V.; Arango, V.; Majo, V. J.; Simpson, N. R.; Heertum, R. V.; Mann, J. J.; Kumar, J. S. D. Bioorg. Med. Chem. 2007, 15, 1802. (b) Uneyama, K. In Organofluorine Chemistry; Blackwell: Oxford, U.K., 2006. (c) Soloshonok, V. A. In Fluorine-Containing Synthons; American Chemical Society: Washington, DC, 2005.

(11) Romanenko, V. D.; Kukhar, V. P. Chem. Rev. 2006, 106, 3868− 3935.

(12) (a) Jakeman, D. L.; Ivory, A. J.; Wiliamson, M. P.; Blackburn, G. M. J. Med. Chem. 1998, 41, 4439−4452. (b) Herczegh, P.; Buxton, T. B.; McPherson, J. C.; Kovács-Kulyassa, A.; Brewer, P. D.; Sztaricskai, F.; Stroebel, G. G.; Plowman, K. M.; Farcasiu, D.; Hartmann, J. F. J. Med. Chem. 2002, 45, 2338−2341. (c) Makhaeva, G. F.; Aksinenko, A. Y.; Sokolov, V. B.; Baskin, I. I.; Palyulin, V. A.; Zefirov, N. S.; Hein, N. D.; Kampf, J. W.; Wijeyesakere, S. J.; Richardson, R. J. Chem. Biol. Interact. 2010, 187, 177−184.

(13) (a) Piras, H.; Fleming, I. N.; Harrison, W. T. A.; Zanda, M. Synlett 2012, 44, 2899−2902. (b) Binkert, C.; Frigerio, M.; Jones, A.; Meyer, S.; Pesenti, C.; Prade, L.; Viani, F.; Zanda, M. ChemBioChem 2006, 7, 181−186. (c) Lazzaro, F.; Crucianelli, M.; De Angelis, F.; Frigerio, M.; Malpezzi, L.; Volonterio, A.; Zanda, M. Tetrahedron: Asymmetry 2004, 15, 889−893. (d) Volonterio, A.; Bellosta, S.; Bravin, F.; Bellucci, M. C.; Bruche, L.; Colombo, G.; Malpezzi, L.; Mazzini, S.; Meille, S. V.; Meli, M.; Ramirez de Arellano, C.; Zanda, M. Chem. Eur. J. 2003, 9, 4510−4522. (e) Molteni, M.; Volonterio, A.; Zanda, M. Org. Lett. 2003, 5, 3887−3890.

(14) Cal, D. Tetrahedron Lett. 2012, 53, 3774−3776.

(15) (a) Röschenthaler, G.-V.; Kukhar, V. P.; Kulik, I. B.; Sorochinsky, A. E.; Soloshonok, V. A. J. Fluorine Chem. 2011, 132, 834−837. (b) Röschenthaler, G. V.; Kukhar, V.; Barten, N.; Gvozdovska, N.; Belik, M.; Sorochinsky, A. Tetrahedron Lett. 2004, 45, 6665−6667. (c) Sergeeva, N. N.; Golubev, A. S.; Henning, L.; Burger, K. Synthesis 2003, 915−919. (d) Nieschalk, J.; Batsanov, A. S.; O'Hagan, D.; Howard, J. Tetrahedron 1996, 52, 165−176.

(16) Palacios, F.; Ochoa de Retana, A. M.; Alonso, J. M. J. Org. Chem. 2006, 71, 6141−6148.

(17) (a) Das Sarma, K.; Zhang, J.; Huang, Y.; Davidson, J. G. Eur. J. Org. Chem. 2006, 3730−3737. (b) Rose, G. M.; Hopper, A.; De Vivo,

The Journal of Organic Chemistry Article and the Second Secon

M.; Tehim, A. Curr. Pharm. Des. 2005, 11, 3329−3334. (c) Shorvon, S. Lancet 2001, 358, 1885−1892. (d) Reddy, P. A.; Hsiang, B. C. H.; Latifi, T. N.; Hill, M. W.; Woodward, K. E.; Rothman, S. M.; Ferrendelli, J. A.; Covey, D. F. J. Med. Chem. 1996, 39, 1898−1906. (e) Kazmierski, W. M.; Andrews, W.; Furfine, E.; Spaltenstein, A.; Wright, L. Bioorg. Med. Chem. Lett. 2004, 14, 5689−5692. (f) Spaltenstein, A.; Almond, M. R.; Bock, W. J.; Cleary, D. G.; Furfine, E. S.; Hazen, R. J.; Kazmierski, W. M.; Salituro, F. G.; Tung, R. D.; Wright, L. L. Bioorg. Med. Chem. Lett. 2000, 10, 1159−1162. (g) Tang, K.; Zhang, J. T. Neurol. Res. 2002, 24, 473−478. (h) Rose, G. M.; Hopper, A.; De Vivo, M.; Tehim, A. Curr. Pharm. Des. 2005, 11, 3329−3334.

(18) (a) Lee, C.; Choi, E.; Cho, M.; Lee, B.; Oh, S. J.; Park, S.-K.; Lee, K.; Kim, H. M.; Han, G. Bioorg. Med. Chem. Lett. 2012, 22, 4189− 4192. (b) Cherney, R. J.; Mo, R.; Meyer, D. T.; Voss, M. E.; Yang, M. G.; Santella, J. B.; Duncia, J. V.; Lo, Y. C.; Yang, G.; Miller, P. B.; Scherle, P. A.; Zhao, Q.; Mandlekar, S.; Cvijic, M. E.; Barrish, J. C.; Decicco, C. P.; Carter, P. H. Bioorg. Med. Chem. Lett. 2010, 20, 2425− 2430.

(19) de los Santos, J. M .; Lopez, Y.; Aparicio, D.; Palacios, F. J. Org. Chem. 2008, 73, 550−557.

(20) Palacios, F.; Ochoa de Retana, A. M.; Pascual, S.; Oyarzabal, J. J. Org. Chem. 2004, 69, 8767−8774.

(21) Vicario, J.; Aparicio, D.; Palacios, F. J. Org. Chem. 2009, 74, 452−455.

(22) (a) Palacios, F.; Alonso, C.; Pagalday, J.; Ochoa de Retana, A. M.; Rubiales, G. Org, Bioorg. Chem 2003, 1, 1112−1118. (b) Barluenga, J.; López, F.; Palacios, F. J. Organomet. Chem. 1990, 382, 61-67.

(23) Palacios, F.; Perez de Heredia, I.; Rubiales, G. J. Org. Chem. 1995, 60, 2384−2390. Formation of the phosphazenes 1 was detected by nitrogen gas formation during the addition of phosphine and was monitored by 31P NMR. The spectra showed the disappearance of the phosphine signal and appearance of a new signal in the range of δ 2− 10 ppm of the corresponding phosphazenes 1.

(24) Palacios, F.; Alonso, C.; Aparicio, D.; Rubiales, G.; De los Santos. Tetrahedron 2007, 63, 523−575.

(25) Tverdomed, S. N.; Kolanowski, J.; Lork, E.; Röschenthaler, G.-V. Tetrahedron 2011, 67, 3887−3903.

(26) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J. A., Jr.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Keith, T.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J. M.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, O.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. Gaussian 09, Revision B.01; Gaussian, Inc., Wallingford, CT, 2010.

(27) (a) Parr, R. G.; Yang, W. In Density-Functional Theory of Atoms and Molecules; Oxford University Press: Oxford, U.K., 1989. (b) Ziegler, T. Chem. Rev. 1991, 91, 651−667.